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VIRTUAL SCREENING AND MOLECULAR DOCKING STUDIES OF SEVERAL PLANT COMPOUNDS WITH THE PRIMARY PROTEASE OF THE COVID-9 VIRUS

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ABSTRACT

Researchers have tried to find a compound that can inhibit the replication of the SARS-CoV-2 virus since the outbreak of the Covid-19 pandemic. The present study evaluates the bioactive compounds found in several plants using a molecular binding approach to inhibit the primary protease of SARS-CoV-2. This study investigated 40 different herbal compounds with the 6Y2F protein of coronavirus. Auto Dock Vina 1.5.6 software was used to evaluate molecular binding. Validation was performed in PyMol software. The results were also analyzed by Biovia Discovery Studio 4.5. The best protein-ligand complex compound was selected by determining the binding score that had the highest affinity (the most negative ΔG Gibbs binding free energy). Among 40 herbal compounds, 21 herbal compounds showed a high energy of -8.0 kJ/mol. Based on the results of binding energy and RMSD value, among the dockings performed, 8 compounds including Ganoderic acid C2, Ursolic Acid, Lupeol, Kuwanon B, Emodin-8-glucoside, Adonitoxin, Kuwanon E, and Isohemiphloin are recommended for further studies in the invivo and invitro sections.

Keywords: Coronavirus, SARS-CoV-2, Herbal compounds, Molecular binding, Auto Dock Vina

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INTRODUCTION

A new respiratory viral disease was reported in Wuhan, China, in December 2019. It was an infectious disease caused by severe acute respiratory syndrome of the SARS-CoV-2 virus [1]. This disease spread worldwide, leading to an ongoing epidemic [2]. Different types of this virus have appeared in many countries until now. The most dangerous types are the alpha, beta, gamma, delta, and omicron strains [3]. About 551 million patients and more than 6 million deaths from this virus were confirmed up to July 2022, making this disease one of the deadliest in history [4]. Although investigations have revealed that various drugs are effective against viruses belonging to the same group, none of them has shown the same potential as a treatment for COVID-19. The primary protease of the COVID-19 virus is considered an attractive target for the study of antiviral drugs against the SARS-CoV-2 virus and other coronavirus infections. Covid-19 symptoms vary but often include fever, cough, headache, fatigue, breathing problems, and loss of smell and taste [5, 6]. This complicated situation has resulted in searching for new treatments and rapid practical measures to treat the disease and reduce its prevalence. Hence, understanding how this virus works and spreads is vital for developing a vaccine.

Further studies are still needed to find effective drugs to inhibit the virus and specific treatment regimens to overcome morbidity and mortality since COVID-19 is a new disease with severe health problems. Covid-19 is very similar to the SARS-CoV-2 virus. There are primary five therapeutic protein targets for SARS-CoV-2, including angiotensin-converting enzyme 2 (ACE2), spike protein, major protease (M pro), RNA-dependent RNA polymerase (RdRp), and papain-like protease. In microscopic imaging, SARS-CoV-2 with its crown-like surface protrusion appears to belong to the family of beta-coronaviruses, which have encapsulated and single-stranded RNA. They primarily infect host lung cells by binding to the angiotensin-converting enzyme 2 (ACE2) receptor [7]. The viral genome is translated as mRNA by the host cell machinery and produces enzymes necessary for RNA synthesis, including RNA-dependent RNA polymerase [8]. Then, these polyproteins are divided into structural proteins such as RdRp, and PL pro. Inhibition can block the synthesis of viral proteins. Thus, it plays a vital role in the viral life cycle [9].

Plants are one of the sources of active medicinal compounds used extensively in the treatment of diseases [10-11]. Many reported bioactive plant compounds have antifungal, antibacterial, and antiviral activities [12]. Plants and plant-derived products have advantages such as simplicity, greater safety, less toxicity, lower cost, higher speed of action, and compatibility with the environment compared to conventional treatment methods [13]. Drugs were extracted from natural sources in the past. These natural sources are still a major source of conductor compounds and new drugs. Nowadays, much attention is paid to pre-studies in drug design using bioinformatics methods to reduce cost and time in drug production. The use of bioinformatics tools and calculation methods that predict the effectiveness of medicinal compounds and their possible toxicity with a high confidence factor has been much considered in recent years [14]. Molecular docking, simulation, determining target point, and chemical stability studies are among the most significant bioinformatics methods used in drug design. In this regard, molecular docking plays a unique role. By considering the different states of the desired molecules in the three-dimensional space and predicting how the protein (receptor) interacts with bioactive compounds (ligand), it is possible in this technique to examine their interactions and the effective factors in the interaction and determine the more stable and important action in terms of drug identification [15].

A computational binding approach using various molecular binding software such as Auto Dock [16] provides the opportunity to identify and evaluate the bindings and efficiency of various inhibitors of natural and synthetic origin. The appropriateness of the drug can be determined by analyzing the medicinal properties after evaluating the efficient inhibitors. Although potential therapeutic agents can only be validated after experimental testing, computational binding could be a gateway to faster development of effective drugs against diseases such as COVID-19. The present study aims to find potential inhibitors of coronavirus among several selected herbal

compounds that have antitussive, antipyretic, anti-viral, anti-inflammatory, antioxidant, etc. impacts using molecular docking studies. It also aims to answer the questions of whether the 6Y2F protease of Covid-19 can be the target of selected herbal compounds in clinical trials and what are the interactions between this protein and the compounds.

METHODS

First, the primary sequence of the 6Y2F protein of Covid-19 was extracted from the PDB database (Table 1). Dimethyl Sulfoxide and *tert*-butyl *N*-[1-[(2-*S*)-3-cyclopropyl-1-oxidanylidene-1-[[[(2-*S*),3-*R*)-3-oxidanyl-4-oxidanylidene-1-[(3-*S*)-2-oxidanylidene-pyrrolidin-3-yl]-4-[(phenylmethyl)amino]butan-2-yl]amino]propan-2-yl]-2-oxidanylidene-pyridin-3-yl]carbamate in 6Y2F protein was removed from the protein using Discovery Studio 4.5 software [17]. The three-dimensional structure of 40 plant compounds that had antiviral, antitussive, antipyretic, and anti-inflammatory effects were extracted from Pubchem and ChemSpider databases. Table 2 shows the characteristics of plant compounds. Then, docking was performed by Auto Dock Vina software [18]. Table 3 presents the Grid Box dimensions and its coordinates for docking operations. Kollman Charges were used to determine the overall load. Docking results were analyzed by Biovia Discovery Studio 4.5 software. The validity of the docking operation was confirmed by RMSD determination in PyMol software.

Table 1- Receptor examined in this study

Receptor	Specifications
<u>6Y2F</u>	Crystal structure (monoclinic form) of the complex resulting from the reaction between SARS-CoV-2 (2019-nCoV) main protease and <i>tert</i> -butyl (1-((<i>S</i>)-1-(((<i>S</i>)-4-(benzylamino)-3,4-dioxo-1-((<i>S</i>)-2-oxopyrrolidin-3-yl)butan-2-yl)amino)-3-cyclopropyl-1-oxopropan-2-yl)-2-oxo-1,2-dihydropyridin-3-yl)carbamate (alpha-ketoamide 13b)[19].

Table 2- Specifications of ligands studied in this study

	Plant Name	Compound Name	Molecular Formula	CID	MW
	Arachis hypogaea	Soyasaponin I	<u>C₄₈H₇₈O₁₈</u>	108898*	943.1
	Water lily	Nupharin A	<u>C₄₁H₃₀O₂₆</u>	8709251*	938.7
	Lady's glove	Digoxin	<u>C₄₁H₆₄O₁₄</u>	2006532*	780.9
	Cissampelos	Warifteine	<u>C₃₆H₃₈Cl₂N₂O₆</u>	170074	665.6
	Forsythiae fructus	Forsythiaside A	<u>C₂₉H₃₆O₁₅</u>	5281773	624.6
	Mint	Hesperidin	<u>C₂₈H₃₄O₁₅</u>	10621	610.6
	Water pepper	Rutoside	<u>C₂₇H₃₀O₁₆</u>	5280805	610.5

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Ginsen radix	Panasenoside	<u>C₂₇H₃₀O₁₆</u>	99861 91	610.5
Ziziphi Spinosae Semen	Spinosin	<u>C₂₈H₃₂O₁₅</u>	15569 2	608.6
Mint	Eriocitrin	<u>C₂₇H₃₂O₁₅</u>	83489	596.5
Eriobotryae folium	-	<u>C₂₇H₃₂O₁₄</u>	10507 459	580.5
Citrus reticulata	Naringin	<u>C₂₇H₃₂O₁₄</u>	44242 8	580.5
Chrysantbemi flos	Isorhoifolin	<u>C₂₇H₃₀O₁₄</u>	98511 81	578.5
Boldo	-	<u>C₁₈H₁₆N₈O₇</u> <u>S₃⁻²</u>	2656	552.6
Pheasant's eye	Adonitoxin	<u>C₂₉H₄₂O₁₀</u>	44183 8	550.6
Ganoderma	Ganoderic acid C2	<u>C₃₀H₄₆O₇</u>	57396 771	518.7
Hedysarum multijugum	-	<u>C₂₃H₂₄O₁₁</u>	46899 140	476.4
Chrysanthemi flos	Thermopsoside	<u>C₂₂H₂₂O₁₁</u>	11294 177	462.4
Currant	-	<u>C₂₂H₁₈O₁₁</u>	65064	458.4
Illicium Difengpi KLB Et KIM	Betulinic Acid	<u>C₃₀H₄₈O₃</u>	64971	456.7
Perilla Frutescens	Ursolic Acid	<u>C₃₀H₄₈O₃</u>	64945	456.7
Spinach	Vitamin K	<u>C₃₁H₄₆O₂</u>	52804 83	450.7
Ginsen radix	Kaempferol	<u>C₂₁H₂₀O₁₁</u>	53187 55	448.4
Eriobotryae folium	Isohemiphloin	<u>C₂₁H₂₂O₁₀</u>	42607 891	434.4
Myrrh	-	<u>C₂₀H₁₈O₁₁</u>	53178 47	434.3
Sennae Folium	Emodin-8- glucoside	<u>C₂₁H₂₀O₁₀</u>	99649	432.4
Fritillaria pallidiflora	Imperialine	<u>C₂₇H₄₃NO₃</u>	44297 7	429.6
Ricinus	Lupeol	<u>C₃₀H₅₀O</u>	25984 6	426.7
Farfarae flos	Taraxasterol	<u>C₃₀H₅₀O</u>	11525 0	426.7
Mori cortex	Kuwanon E	<u>C₂₅H₂₈O₆</u>	64404 08	424.5

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Gossampini flos	Mangiferin	<u>C₁₉H₁₈O₁₁</u>	52816 47	422.3
Mori cortex	Kuwanon B	<u>C₂₅H₂₄O₆</u>	44258 295	420.5
zingiberis	beta-sitosterol	<u>C₂₉H₅₀O</u>	22228 4	414.7
Soja semen nigrum	Stigmasterin	<u>C₂₉H₄₈O</u>	52807 94	412.7
Pennyroyal	Cleomiscosin A	<u>C₂₀H₁₈O₈</u>	44251 0	386.4
Elder	(+)-Bicuculline	<u>C₂₀H₁₇NO₆</u>	10237	367.4
Tripterygii radix	Triptolide	<u>C₂₀H₂₄O₆</u>	10798 5	360.4
Viper's-buglosses	-	<u>C₂₀H₂₂O₆</u>	12309 637	358.4
Papaveris pericarpium	-	<u>C₂₂H₂₉NO₃</u>	21287 385	355.5
Knotweed	-	<u>C₂₂H₂₀O₄</u>	11163 864	348.4

*The codes are chemspider.

Table 3- The dimensions and size of the studied protein space

	center_x	center_y	center_z	size_x	size_y	size_z
<u>6Y2F</u>	-4.73	-2.885	12.052	126	126	126

RESULTS

Among the 40 dockings performed, 21 plant compounds with the 6Y2F receptor of the coronavirus have energy above -8.0. Table 4 shows the results whose minimum free energy is less than -8.0 Kcal/mol. It also presents the target proteins and minimum free energy changes (ΔG) and amino acids involved in hydrogen bonding and the RMSD value of each.

Table 4- Interactions and energy results of complexes higher than $\Delta G = -8.0$

Plant Name		ΔG (Kcal/mol)	H - Bond	RMSD (A°)
Lady's glove	Digoxin	-9.6	TYR A:239_LEU A:271_LEU A:272_LYS A:137	3.463
Cissampelos	Warifteine	-9.3	-	0.001
Mint	Hesperidin	-9.1	PHE A:140_CYS A:145_HIS A:163	1.326

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	Chrysanthemi flos	Isorhoifolin	-9.1	PHE A:140_GLU A:166_LEU A:167_GLN A:192_CYS A:145	0.729
	Farfarae flos	Taraxasterol	-9.1	-	0.001
	Arachis hypogaea	Soyasaponin I	-9.0	ASN A:238_LYS A:137_THR A:169_ASN A:133	1.415
	water lily	Nupharin A	-9.0	GLN A:189_MET A:49_VAL A:186_ARG A:188_GLY A:143	1.897
	Mori cortex	Kuwanon E	-8.8	ILE A:152_GLN A:110	1.117
	Chrysanthemi flos	Thermopsoside	-8.7	HIS A:163_CYS A:145_PHE A:140	1.386
	Ganoderma	Ganoderic acid C2	-8.6	ASN A:203_THR A:111_ASN A:151	0.000
	Mint	Eriocitrin	-8.5	GLU A:166_LEU A:167_THR A:190_CYS A:145	2.846
	Citrus reticulata	Naringin	-8.5	LEU A:271_ASN A:238_LEU A:287_ASP A:197_ARG A:131_ASP A:289	2.797
	Perilla Frutescens	Ursolic Acid	-8.4	ASP A:289_AA:131	0.000
	Ricinus	Lupeol	-8.4	ASN A:203	0.000
	Elder	(+)-Bicuculline	-8.4	ASN A:151_GLN A:110	1.928
	Mori cortex	Kuwanon B	-8.3	ASN A:238_TYR A:239_ASP A:197_THR A:198_ARG A:131	0.113
	Forsythiae fructus	Forsythiaside A	-8.2	ASN A:238_LYS A:137_TYR A:239_LEU A:287_THR A:199	1.260

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	Sennae Folium	Emodin-8-glucoside	-8.2	CYS A:145_GLN A:192	0.386
	zingiberis	beta-sitosterol	-8.2	-	0.000
	Eriobotryae folium	Isohemiphloin	-8.1	CYS A:145_GLN A:189	1.137
	Pheasant's eye	Adonitoxin	-8.0	THR A:198_LYS A:137_MET A:276_ALA A:285_LEU A:271	0.063

A minimum number of hydrogen bonds must be formed for the pseudo-drug to affect the receptor [20] and the RMSD value is used to confirm the binding protocol. Thus, results without hydrogen bonding and RMSD values above 1.5 angstroms in Table 4 were omitted, despite having the highest $G\Delta$ values. Table 4 presents 21 compounds. Seven compounds among them showed the best RMSD results, indicating the reliability of the data of this study. Digoxin compound in binding with 6Y2F receptor did not have good RMSD value despite the best binding energy. Ganoderic acid C2, Ursolic Acid, and Lupeol compounds caused protein instability by binding to 6Y2F, which can be seen in RMSD analysis. The RMSD obtained from this compound is zero A° (Figures 1, 2, and 3), indicating the validation of the binding protocol.

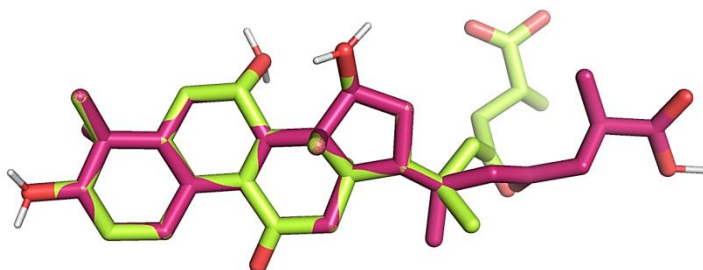


Figure 1- RMSD of 0.0 A° for Ganoderic acid C2 compound of Ganoderic acid C2 by PyMol software

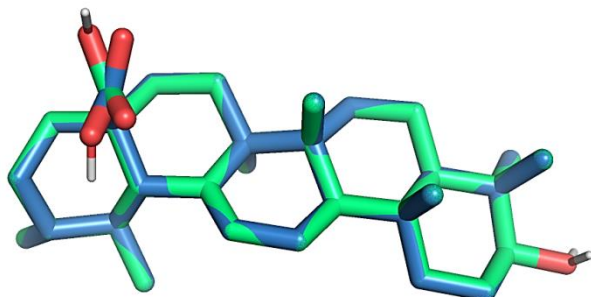


Figure 2- RMSD of 0.0 A° for Ursolic Acid compound of Perilla Frutescens by PyMol software

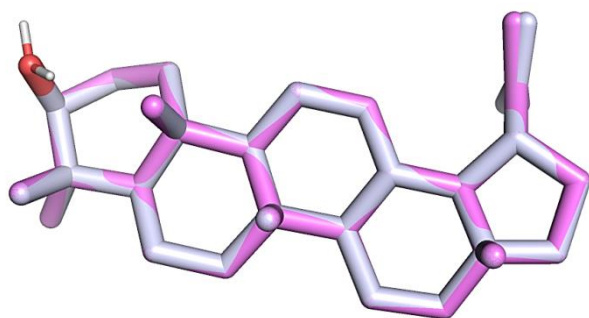


Figure 3- RMSD of 0.0 Å° for lupeol compound by Pymol software

Glycoside compounds such as flavonol glycoside have therapeutic properties including treating headache, and fever, and helping to treat cough, bronchitis, and infectious diseases [21]. The binding scores of glycosidic compounds of digoxin, eriocitrin, hesperidin, naringin, Adonitoxin, and Emodin-8-glucoside with 6Y2F protein were -6.9, -5.8, -1.9, -5.8, -0.8, and -2.8, respectively. The results of a study by Otomo et al. [22], Chen et al. [23], and Adam et al. [24] confirm the results of Hesperidin and Naringin. Based on Lin et al.'s experiments, Emodin-8-glucoside compound has very good water solubility and shows almost no cytotoxicity [25]. Animals that consume plants containing cardiac glycosides, including adonitoxin, usually suffer from fatal digestive and cardiac disorders despite the antioxidant, antimicrobial, anti-inflammatory, cardioprotective, neuroprotective, and antiallergic properties of glycosidic compounds [26]. However, recent studies indicate that using a low dose of this compound has therapeutic properties. In short, phytochemical and pharmacological studies of the Adonis L. genus have received much attention [26-27]. Extracts enriched in cardiac glycosides have been made and the active compounds have been isolated and proven to provide cardioprotective activity. However, plants of this genus should be further investigated and developed with special attention to resource conservation and clinical trials.

Terpenoids are active compounds found in plants. The binding score of Soyasaponin I against the original protease was -9.0. This compound has anti-inflammatory activity in addition to being known as an anti-herpes simplex virus [28] and [29]. However, some studies indicate that it can increase the pathogenicity of the virus to the host [30-31]. Warifteine and Nupharin A are alkaloid compounds with binding scores of -9.3 and -9.0, respectively. They have the properties to treat asthma, inflammatory disorders, bronchitis, antiplatelet, and anticoagulants [32]. Several different classes of bioactive molecules isolated from many plants have antiviral activity [33-34]. One of the methods in determining whether a compound has medicinal potential is to follow Lipinski's rule of five (RO5) [35]. Based on this rule, orally active drugs should not violate more than one of the established criteria [36]. Thus, it was examined whether each docking matched Lipinski's RO5 (Table 5).

Table 5- Review of Lipinski's rules

Compound Name	mass	hydrogen bond donor	hydrogen bond acceptors	Log P
Isorhoifolin	556	8	14	0.85
Ganoderic acid C2	476	4	7	0.0
Ursolic Acid	410	2	3	1.74
Lupeol	337	1	1	0.0
Kuwanon B	399	3	6	0.0
Emodin-8-glucoside	418	6	10	-0.67

Adonitoxin	513	5	10	0.0
Hesperidin	584	8	15	1.093
Soyasaponin I	875	11	18	1.99
Kuwanon E	400	4	6	0.0
Thermoposide	446	6	11	-0.28
Forsythiaside A	597	9	15	-0.34
Isohemiphloin	419	7	10	-3.0

CONCLUSION

This study revealed that the natural compounds Digoxin and Warifteine among the selected plant compounds have better binding free energies with the 6Y2F protein of SARS-CoV-2. Although the molecular binding results of Ganoderic acid C2, Ursolic Acid, Lupeol, Kuwanon B, Emodin-8-glucoside, Adonitoxin, Kuwanon E, and Isohemiphloin are lower than the first two compounds, the analysis of RMSD parameters, interactions, number of hydrogen bonds, and RO5 criteria and their non-toxic properties showed better performance. These compounds have a better potential as antiviral plant chemicals and to solve respiratory, inflammatory, infectious, and coagulation problems, which may prevent the proliferation of the virus or help to treat this disease. These 9 inhibitors are appropriate candidates as drugs for inhibiting the activity of the primary enzyme of the SARS-CoV-2 coronavirus for clinical and laboratory studies. However, the conducted studies are theoretical. Experimental work is required to ensure the accuracy of the data, and the results of this research alone cannot claim that the introduced compounds can inhibit the COVID-19 protease.

REFERENCES

1. Sherif, Y.E., et al.(2021), Phytochemicals of Rhus spp. as potential inhibitors of the SARS-CoV-2 main protease: molecular docking and drug-likeness study. Evidence-Based Complementary and Alternative Medicine, 2021.
2. Zimmer, C.(2021), The Secret Life of a Coronavirus–An oily, 100-nanometer-wide bubble of genes has killed more than two million people and reshaped the world. Scientists don't quite know what to make of it. Scientists don't quite know what to make of it. Retrieved, 28.
3. Cedro-Tanda, A ,et al.(2021), The evolutionary landscape of SARS-CoV-2 variant B. 1.1. 519 and its clinical impact in Mexico City. Viruses, 13(11): p. 2182.
4. COVID, G. (2021), database Retrieved from <https://github.com/CSSEGISandData/COVID-19>.
5. Saniasiaya, J. (2019), M.A. Islam, and B. Abdullah, Prevalence of olfactory dysfunction in coronavirus disease (COVID-19): a meta-analysis of 27,492 patients. The Laryngoscope, 2021. 131(4): p. 865-878.
6. Agyeman, A.A. (2020), et al. Smell and taste dysfunction in patients with COVID-19: a systematic review and meta-analysis. in Mayo Clinic Proceedings.. Elsevier.
7. Li, W., et al. (2003), Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. Nature, 426(6965): p. 450-454.
8. Poduri, R., G. Joshi, and G. Jagadeesh, (2020), Drugs targeting various stages of the SARS-CoV-2 life cycle: Exploring promising drugs for the treatment of Covid-19. Cellular signalling,. 74: p. 109721.
9. Rathnayake, A.D., et al. (2020), 3C-like protease inhibitors block coronavirus replication in vitro and improve survival in MERS-CoV–infected mice. Science translational medicine, 12(557): p. eabc5332.

10. Shakeran, Z., M. Nosrati, and Z. Shakeran. (2018), In silico screening of hepatitis C virus NS3/4A protease inhibitor (s) from *Cornus officinalis* and *Syzygium aromaticum*.
11. Shakeran, Z., M. Nosrati, and Z. Shakeran. (2018), In silico Screening of Hepatitis C Virus NS3/4A Protease Inhibitor (s) from medicinal plants. *Razi Journal of Medical Sciences*, 25(167): p. 69-80.
12. Cheng, P.W., et al. (2006), Antiviral effects of saikosaponins on human coronavirus 229E in vitro. *Clinical and Experimental Pharmacology and Physiology*, 33(7): p. 612-616.
13. Ameer, K., H.M. Shahbaz, and J.H. (2017), Kwon, Green extraction methods for polyphenols from plant matrices and their byproducts: A review. *Comprehensive Reviews in Food Science and Food Safety*, 16(2): p. 295-315.
14. Nantasenamat, C., C. Isarankura-Na-Ayudhya, and V. Prachayasittikul. (2010), Advances in computational methods to predict the biological activity of compounds. *Expert opinion on drug discovery*, 5(7): p. 633-654.
15. Ferreira, L.G., et al. (2015), Molecular docking and structure-based drug design strategies. *Molecules*, 20(7): p. 13384-13421.
16. Morris, G.M., et al. (2001), AutoDock. Automated docking of flexible ligands to receptor-User Guide.
17. Studio, D.(2015), Dassault systemes BIOVIA, Discovery studio modelling environment, Release 4.5. Accelrys Softw Inc, 2015: p. 98-104.
18. Huey, R., G.M. Morris, and S. Forli. (2012), Using AutoDock 4 and AutoDock vina with AutoDockTools: a tutorial. The Scripps Research Institute Molecular Graphics Laboratory, 1055 : p. 92037.
19. Zhang, L., Lin, D., Sun, X., Curth, U., Drosten, C., Sauerhering, L., ... & Hilgenfeld, R. (2020). Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved α -ketoamide inhibitors. *Science*, 368(6489), 409-412.
20. Chen, D., et al. (2016), Regulation of protein-ligand binding affinity by hydrogen bond pairing. *Science advances*, 2(3): p. e1501240.
21. Brett, G.M., et al. (2008), Absorption, metabolism and excretion of flavanones from single portions of orange fruit and juice and effects of anthropometric variables and contraceptive pill use on flavanone excretion. *British Journal of Nutrition*, 101(5): p. 664-675.
22. Utomo, R.Y., M. Ikawati, and E. (2020), Meiyanto, Revealing the potency of citrus and galangal constituents to halt SARS-CoV-2 infection.
23. Chen, Y.W., C.-P.B. Yiu, and K.-Y.(2020), Wong, Prediction of the SARS-CoV-2 (2019-nCoV) 3C-like protease (3CL pro) structure: virtual screening reveals velpatasvir, ledipasvir, and other drug repurposing candidates. *F1000Research*, 9.
24. Adem, S., et al.(2020), Identification of potent COVID-19 main protease (Mpro) inhibitors from natural polyphenols: an in silico strategy unveils a hope against CORONA.
25. Lin, H.-W., et al.(2010), Anti-HIV activities of the compounds isolated from *Polygonum cuspidatum* and *Polygonum multiflorum*. *Planta medica*, 76(09): p. 889-892.
26. Galey, F.D., et al.(1996), Diagnosis of oleander poisoning in livestock. *Journal of Veterinary Diagnostic Investigation*, 8(3): p. 358-364.
27. Zhang, L.-h., et al.(2015), Determination of other related carotenoids substances in astaxanthin crystals extracted from *Adonis amurensis*. *Journal of Oleo Science*, 64(7): p. 751-759.
28. Shang, X., et al.(2019), The genus *Adonis* as an important cardiac folk medicine: a review of the ethnobotany, phytochemistry and pharmacology. *Frontiers in Pharmacology*, 10: p. 25.
29. Blevins, R.D. and M.P. (1980), Domic, The effect of Δ -9-tetrahydrocannabinol on herpes simplex virus replication. *Journal of General Virology*, 49(2): p. 427-431.
30. Reiss, C.S. (2010), Cannabinoids and viral infections. *Pharmaceuticals*, 3(6): p. 1873-1886.
31. Tahamtan, A., et al. (2018), Effects of cannabinoid receptor type 2 in respiratory syncytial virus infection in human subjects and mice. *Virulence*, 9(1): p. 217-230.
32. Roberts, M.F.(2013), Alkaloids: biochemistry, ecology, and medicinal applications.

Springer Science & Business Media.

33. Ullah, S., et al. (2022), Identification of phytochemical inhibitors of SARS-CoV-2 protease 3CLpro from selected medicinal plants as per molecular docking, bond energies and amino acid binding energies. *Saudi Journal of Biological Sciences*, 29(6): p. 103274.
34. Mukhtar, M., et al. (2008), Antiviral potentials of medicinal plants. *Virus research*, 131(2): p. 111-120.
35. Lipinski, C.A., et al. (1997), Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Advanced drug delivery reviews*, 23(1-3): p. 3-25.
36. Denaro, M., et al. (2020), Antiviral activity of plants and their isolated bioactive compounds: An update. *Phytotherapy Research*, 34(4): p. 742-768.